

## A PENTACYCLIC TRITERPENE DIACID FROM *MYRIANTHUS ARBOREUS*

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**Key Word Index**—*Myrianthus arboreus*; Cecropiaceae; bark; euscaphic acid; tormentic acid; myrianthinic acid; arboreic acid; myriaboric acid.

**Abstract**—In addition to myrianthinic acid, arboreic acid, and the known tormentic and euscaphic acids, a new ursane derivative, myriaboric acid, has been isolated from the polar material of the stem bark of *Myrianthus arboreus*, and its structure elucidated by spectroscopic methods.

### INTRODUCTION

*Myrianthus arboreus* P. Beauv. [1] is a small tropical African tree, which grows from Guinea to Uganda, through Angola and Tanzania. Previous works on *M. arboreus* described the isolation of peptide alkaloids [2], and of many triterpene acids [3-5]. Recently, we obtained two novel pentacyclic triterpenoids, myrianthinic acid [6] and arboreic acid [7] from the polar extract of the plant, both as their methyl esters. We now report, from the same source, the structural elucidation of a new pentacyclic diacid, myriaboric acid, as its dimethyl ester derivative (1).

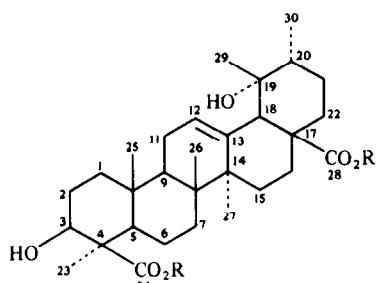
### RESULTS AND DISCUSSION

The methylated ethyl acetate extract of pulverized bark gave the methyl ester of myriaboric acid (1), which crystallized from hexane as colourless needles, mp 258-260°. A positive response to Liebermann-Buchard, TCA [8] and TNM tests showed compound 1 to be an unsaturated pentacyclic triterpene. Its molecular formula was established as  $C_{32}H_{50}O_6$  ( $M^+$  at  $m/z$  530). The IR spectrum of 1 exhibited main absorptions at  $\nu_{max}$  3510 and 3460 (sharp, OH), 1725, 1710 (-OAc), 1630 (trisubstituted double bond), 1050, 1000 (secondary OH) and 930 (tertiary OH)  $cm^{-1}$  [8]. The 200 MHz  $^1H$  NMR spectrum of 1 in  $CDCl_3$  showed two carbomethoxyl signals at  $\delta$  3.60 and 3.76, a vinylic proton at  $\delta$  5.34, four tertiary methyl groups at  $\delta$  0.71 (3H, Me-26), 0.89 (3H, Me-29), 1.13 (3H, Me-25), 1.22 (3H, Me-27), 1.30 (3H, Me-23) and a secondary methyl group at  $\delta$  0.94 (3H, d,  $J$  = 6.9 Hz, Me-30) [9].

Since the two carbomethoxyl groups accounted for four of the six oxygens present in 1, the remaining oxygens should exist as hydroxyl functions if an ursane skeleton is assumed for the compound.

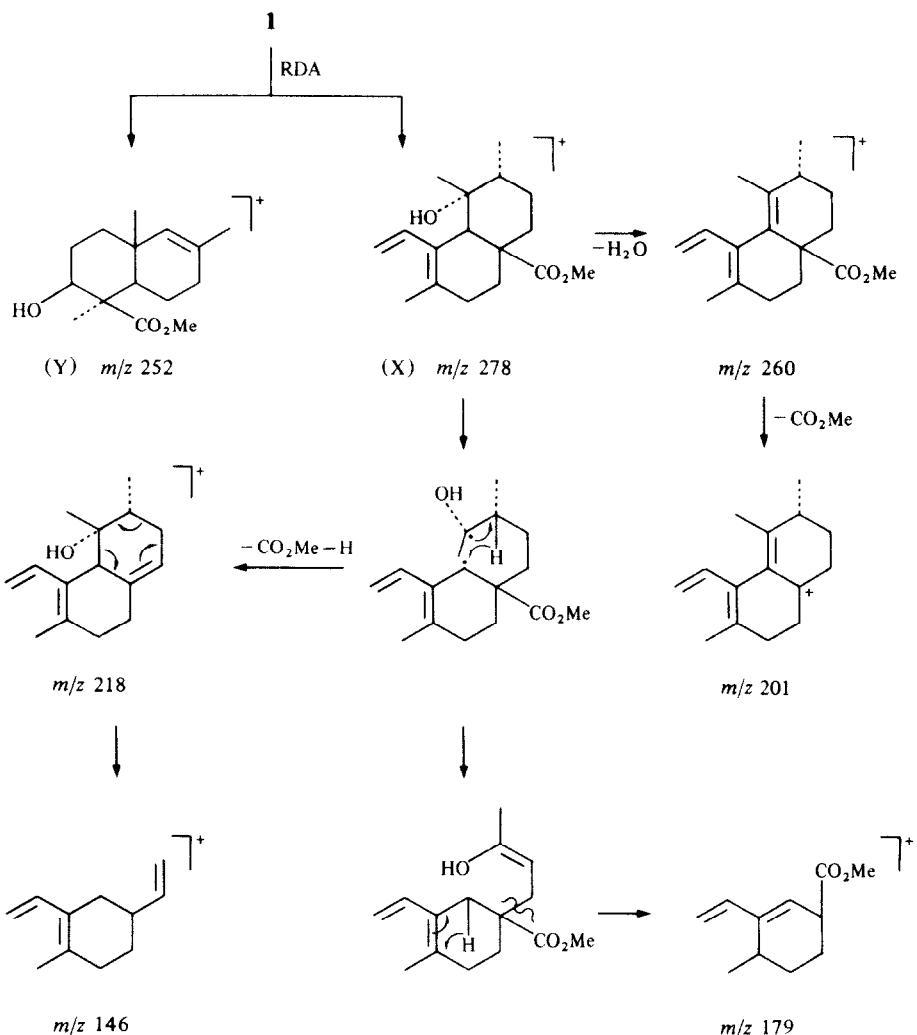
The mass spectrum of 1 indicated that by a retro-Diels-Alder fragmentation of ring C, two main fragments [X,  $m/z$  278 and Y,  $m/z$  252 (Scheme)] were produced, which were quite diagnostic. The presence in 1 of four tertiary methyl groups, one secondary methyl group, two carbomethoxyl groups and the chemical shift of H-18 [9] in conjunction with the mass spectral fragmentation [10]

confirmed that the compound was an urs-12-ene derivative, carrying a hydroxyl and a carbomethoxyl both in ring A/B and in ring D/E. The fragment obtained from the retro-Diels-Alder rupture of ring C at  $m/z$  278, and subsequent fragmentations from the latter ion suggested the same ring D/E substitution as in methyltormentate 4 [11]. Moreover, the multiplicity of the hydrogen atom H-18 was in accord with the presence of a substituent on C-19. The above proton resonance which appeared as a singlet at  $\delta$  2.59, agreed with the existence of a 19 $\alpha$ -hydroxyl group. Also, the presence of the hydroxyl was corroborated by a downfield effect on Me-29, which shifted from  $\delta$  0.80 to 0.93, as it does in methyl urs-12-en-28-Oate [4]. The hydroxyl in ring A/B was assigned to the 3 $\beta$ -position, on biogenetic grounds and because of the presence of a 3 $\alpha$  proton at  $\delta$  3.20, as a multiplet on its  $^1H$  NMR spectrum [9]. The carbomethoxyl in ring A/B had to be located at C-23, C-24, C-25, or C-26. Two main arguments led us to locate the carbomethoxyl at C-24: from Bory and co-workers [12], when a di or a triterpene carries an equatorial carbomethoxyl function, a very intense IR absorption band is observed at  $\nu_{max}$  1245  $\pm$  5  $cm^{-1}$ . But when the carbomethoxyl is axially oriented, three characteristic bands are observed at  $\nu_{max}$  1155  $\pm$  4, 1190  $\pm$  5 and 1230  $\pm$  5  $cm^{-1}$ . In the IR spectrum of 1, three intense absorption bands were observed at  $\nu_{max}$  1159, 1185 and 1235  $cm^{-1}$ . This result is in good agreement with the axial orientation of the two carbomethoxyl



1 R = Me

2 R = H



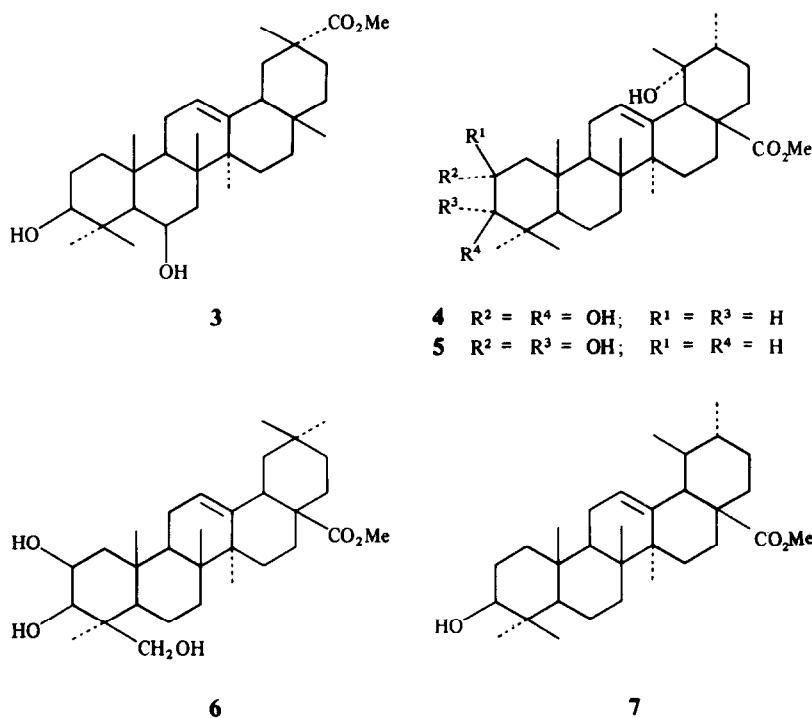
groups in the molecule, and consequently, the ester in ring A/B is also axially oriented. The second argument in favour of the location of the above carbomethoxyl at C-24 is the downfield effect on the C-23 methyl group which appeared at  $\delta$ 1.30 in compound 1, compared to its position at  $\delta$ 0.96 in methyl urs-12-en-28-Oate [9, 13, 14]. Accordingly, the carbomethoxyl was finally located at C-24.

The  $^{13}\text{C}$  NMR spectrum strongly supported structure 1. Close inspection of the spectrum led us to rule out most of the known triterpene skeletons and to retain that of an ursane derivative [15]. The signals were attributed by means of single frequency off-resonance decoupling techniques [16,17] and application of known chemical shift rules involving different substitution shifts, and steric effects [17], as well as by comparison with  $^{14}\text{C}$  NMR spectral data of known ursane derivatives [15, 18]. More specifically, comparison of the  $^{13}\text{C}$  NMR spectrum of compound 1 with that of  $\alpha$ -amyrin (7) was especially helpful (Table 1). The C-18 carbon resonated at  $\delta$ 53.7 in compound 1 and  $\delta$ 58.9 in 7, this difference of up to 4 ppm resulted in an upfield effect due to the C-28 carbomethoxyl group in compound 1 [19]. Similarly, C-22 resonated at  $\delta$ 37.4 in 1 and  $\delta$ 41.5 in 7, for the same reason. An analogous effect was observed for C-3 which appeared at  $\delta$ 73.2 in 1 and 78.8 in  $\alpha$ -amyrin, because of

Table 1.  $^{13}\text{C}$  NMR data for compounds 1 and 7 [15]

C	1	7	C	1	7
1	41.3	38.7	17	48.5	33.7
2	27.5	27.2	18	53.7	58.9
3	73.2	78.8	19	77.6	39.6
4	48.1	38.7	20	41.1	39.6
5	53.1	55.2	21	25.7	31.2
6	20.9	18.3	22	37.4	41.5
7	33.3	32.9	23	25.5	28.1
8	41.3	40.0	24	177.0	15.6
9	47.1	47.7	25	15.7	15.6
10	41.5	36.9	26	17.3	16.8
11	25.1	17.4	27	26.1	23.3
12	129.1	124.3	28	178.2	28.1
13	138.6	139.3	29	27.4	23.3
14	41.3	42.0	30	16.1	21.3
15	28.6	28.7	24CO <sub>2</sub> Me	52.1	
16	25.5	26.6	28CO <sub>2</sub> Me	51.5	

the presence of C-24 carbomethoxyl group. The carbonyl of the two carbomethoxyls resonated at  $\delta$ 177.0 and 178.2 for C-24 and C-28 respectively. These values agreed with those observed in the same positions in other derivatives



[18]. On the other hand, the carbons  $\alpha$  to these carbo-methoxyls were subjected to a downfield effect. In fact, C-4 and C-17 appeared at  $\delta$  48.1 and 48.5 respectively in 1 and  $\delta$  38.7 and 33.7 respectively in compound 7. These chemical shifts agree with the proposed structure. In view of the above evidence, myriaboric acid methyl ester was assigned structure 1 and accordingly, myriaboric acid is  $3\beta,19\alpha$ -dihydroxyurs-12-en, 24,28-dioic acid (2), which to our knowledge, has not yet been reported in the literature.

During the course of this investigation, myrianthinic acid, arboreic acid, both recently reported [6,7] and the known tormentic and euscaphic acids were also isolated, all as their methyl esters 3,6,4, and 5 respectively.

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#### REFERENCES

1. Berg, C. (1978) *Taxon* **27**, 39
2. Marchand, J., Monseur, X. and Pais, M. (1968) *Ann. Pharm. Fr.* **26**, 771
3. Ojinnaka, C. M., Okogun, J. I. and Okorie, D. A. (1980) *Phytochemistry* **19**, 2482
4. Ojinnaka, C. M., Okogun, J. I. and Okorie, D. A. (1984) *Phytochemistry* **23**, 1127.
5. Ojinnaka, C. M. (1985) *J. Nat. Prod.* **48**, 1002.
6. Ngounou, F. N., Lontsi, D. and Sondengam, B. L. (1988) *Phytochemistry* **27**, 301.
7. Ngounou, F. N., Lontsi, D., Ayafor, J. F. and Sondengam, B. L. (1987) *Phytochemistry* **26**, 3080.
8. Takahashi, K., Kawashi, S., Nishimura, K. I., Kubota, K., Tanabe, Y. and Takani, M. (1974) *Chem. Pharm. Bull.* **22**, 650
9. Cheung, H. T. and Williamson, D. G. (1969) *Tetrahedron* **25**, 119.
10. Karliner, J. and Djerassi, C. (1966) *J. Org. Chem.* **31**, 1945.
11. Potier, P., Das, D. C., Bui, A.-M., Janot, M.-M., Pourrat, A. and Pourrat, H. (1966) *Bull. Soc. Chim. Fr.* **11**, 3458.
12. Bory, S. and Fetizon, M. (1964) *Bull. Soc. Chim. Fr.* **352**.
13. Tursch, B., Savoir, R. and Chiurdoglu, G. (1966) *Bull. Soc. Chim. Belges* **75**, 107.
14. Tursch, B., Savior, R., Ottinger, R. and Chiurdoglu, G. (1967) *Tetrahedron Letters* 539.
15. Seo, S., Tomita, Y. and Tori, K. (1975) *Tetrahedron Letters* 7.
16. Tomita, Y. and Seo, S. (1973) *J. Chem. Soc. Chem. Comm.* 707.
17. Wehrlin, F. W. and Wirthlin, T. (1980) in *Interpretation of Carbon-13 NMR Spectra*. Heyden, London.
18. Miana, G. A. and Hazimi, M. G.-Al (1986) in *Essays on Science*, p. 171. Hamdard Foundation Press, Pakistan.
19. Chen, T. K., Ales, D. C., Baenziger, N. C. and Wiemer, D. F. (1983) *J. Org. Chem.* **48**, 3525.